AMENDMENTS TO THE SPECIFICATION:

Page 15, replace the paragraph beginning on line 12 and ending on line 17 and replace it with the following amended paragraph and table:

--The P60R5NEW primer (5'TCGGATCCTCTAGATCTCTCTG 3') (SEQ ID NO: 5) was used for the amplification of different portions of the region upstream of the *AtMYB60* gene, in combination with the primers reported in the following Table. A BamHI restriction site (GGATCC) was introduced in the P60R5NEW primer.--

Primer	Sequence 5'-3'	Region amplified with P60F1	Construct
P60F1	AAGCTTCACAAGGACACAAGGACA (SEQ ID NO: 6)	1291bp	*p1.3:GUS* *p1.3-2.2:GUS*
P60F8	ATAGAATCTAACACTACTAATTGTTAT (SEQ ID NO: 7)	999bp	*p.09:GFP*
P60F2bis	AAGCTTCAAGTTGCAGTGAATGA (SEQ ID NO: 8)	603bp	*p0.6:GUS*
P60F3	AAGCTTCGTGTGGAGATCAACAT (SEQ ID NO: 9)	246bp	*p0.2:GUS*
P60F5	AAGCTTGCAGAGTGACTCGTGA (SEQ ID NO: 10)	189 bp	*P189:GUS*

Page 15, replace the paragraph beginning on line 20 and bridging pages 15 and 16 with the following amended paragraph:

--The 3' genomic region, 2219bp in length, was amplified using the primers 60-3'UTRF2 (5' CACTTGATGGAGCTCTCTAATATG 3') (SEQ ID NO: 11) and 60-3' UTRR1 (5' CTGCAGACGTTTGTCTAGTAG 3') (SEQ ID NO: 12).--

Page 19, replace the paragraph beginning on line 9 with the following amended paragraph:

--After phenol chloroform extraction, the RNA was precipitated at 4°C in 4M LiCI, washed with 70% ethanol and resuspended in water treated with diethylpyrocarbonate (1% DEPC). 5µg total RNA were treated for 30 min with DnaseI (15 units-Boheringer Mannheim), following the manufacturer's protocol. The reverse-transcription reaction was performed with Reverse Transcriptase Superscript™ II (Life Technologies), according to the manufacturer's indications, using the oligo(dT) primer, formed by 17 dT residues and by the adapter 5'-GGGAATTCGTCGACAAGC-3' (SEQ ID NO: 19). The cDNA samples were amplified in a reaction mixture containing Red Taq PCR Reaction Buffer 1X (Sigma) and 5mM dATP, dCTP, dGTP and dTTP, 25 µM specific primers (Table below), 1 unit RED Taq ™ polymerase (Sigma) and sterile distilled water to a final volume of 25 µl. The amplification was carried out under the following conditions: 1 min at 94°C; 20 cycles at 94°C for 15 sec, 60°C for 15 sec, 72°C for 1 min; 72°C for 10 min. The PCR products were separated on 1% agarose gel and transferred to Hybond N+ filters (Amersham) in 0.4N NaOH. Filters were hybridized with TSB1-, GUS- or GFP-

specific probes amplified using the primers indicated in the Table below, and tagged with digoxigenin using the DIG-High Prime kit (Roche), following the manufacturer's instructions.--

Page 20, replace the table beginning on line 1 with the following amended table:

--Primer	Gene	Sequence 5'-3' (SEQ ID NOS 13-18 respectively in order of appearance)
TSFB1	TSB1	5'-CTCATGGCCGCCGGATCTTGA-3'
TSBR1	TSB1	5'-CTTGTCTCTCCATATCTTGAGCA-3'
GFPF1	GFP	5'-GGAGAAGAACTTTTCACTGGAGTTGTCCC-3'
GFPR1	GFP	5'-TAGTTCATCCATGCCATGTGTAATCCCAGC-3'
GUSF1	GUS	5'-AATAACGGTTCAGGCACAGC-3'
GUSR1	GUS	5'-CTGTGGAATTGATCAGCGTTG-3'--